

*Note: for laboratory research use only*

# **M13 Phage DNA Rapid Extraction Kit (Spin-column)**

*A kit for the isolation of single strand DNA from M13 phage*

**Cat. #: DP2301 (50 preps)**

## I.Kit Content, Storage and Stability

Content	Storage	50 preps (DP2301)
Buffer MB	RT	25 ml
Buffer WB	RT	15 ml
		<i>Add 60ml ethanol before use</i>
Buffer EB	RT	10 ml
Spin-column AC	RT	50
Collection Tube (2ml)	RT	50

*All reagents are stable for 18 months at RT if stored properly.*

### Reminder:

- 1 · *Add ethanol to Buffer WB before use, mix adequately, and then check the box on the label showing ethanol has been added!*
- 2 · Buffer MB may precipitate under low temperature. Incubate to 37°C for a few minutes **until buffer is clear**, and then let the buffer cool to RT before use.
- 3 · Keep the reagent lids tightly capped when not in use to prevent evaporation, oxidation, and changes in pH.

## II .Principle:

The infected bacterial culture is centrifuged to pellet the bacterial cells, and high-salt buffer is added to the supernatant. Phage DNA is selectively adsorbed to silica membrane in a spin-column. Contaminants such as proteins and polysaccharides are efficiently removed by two wash steps. Pure phage DNA is eluted in buffer EB or DNase-free water.

### III. Features:

- 1 · Rapid and convenient. Isolation can be finished in 10 minutes.
- 2 · The kit does not contain poisonous phenol. No need for ethanol precipitation. Multi-elution can ensure highly-purified DNA, which can be applied to all kinds of molecular biology experiments, such as PCR, Southern-blot, restriction enzyme digestion, and mammalian cell transfections.
- 3 · Excellent yield (up to 3µg/800µl culture).

### IV. Notes

*Please read this section before your experiment.*

1. All the centrifugation can be performed at room temperature.
2. Buffer MB contains a corrosive compound; please wear latex gloves to avoid contact with skin, eyes, and cloth. **If contact occurs, wash with water or physiological saline.**
3. Purified DNA typically has A260/A280 ratio between 1.7 and 1.9, which is suitable for sequencing and other biological applications.
4. Set a water bath to 60-70°C before use.

### V. Procedure

#### Before Starting

- *Dilute Buffer WB with 60 ml absolute ethanol, vortex adequately, then the check the box on the bottle to indicate the ethanol has been added!*

1. Collect the liquid culture of bacteria infected with M13 phage or phagemid (M13 origin) into 1.5 ml clean tubes, centrifuge at 12,000 rpm for 5 minutes.
2. Transfer 800 µl supernatant to a new clean tube, and add 400 µl Buffer MB and mix thoroughly.

**If the volume of supernatant is more or less than 800 µl, the dosage of Buffer MB should be accordingly increased or reduced.**

3. Transfer the mixture from step 2 into a spin-column AC (place the spin column to a collection tube), centrifuge at 10,000 rpm for 15 minutes, and discard flow-through.

**The spin-column can accommodate 700  $\mu$ l mixture. Please repeat step 3 until all is loaded.**

4. Add 700  $\mu$ l buffer WB (**please check if ethanol is added!**). Centrifuge at 12,000 rpm for 30 seconds and discard flow-through.
5. Place spin-column AC back to the collection tube. Centrifuge at 12,000 rpm for 1 minute. Remove the residue ethanol as much as possible.
6. Transfer spin column AC to a new clean centrifuge tube. Add 60  $\mu$ l buffer EB (preheated in a water bath with 65-70°C) directly onto the silica-membrane. Incubate 1 minute at room temperature. Centrifuge at 13,000 rpm at 1 minute.

**The volume of elution buffer could be adjusted according to needs.**

**Appropriately reduce elution volume can increase concentration. The minimum volume is 50  $\mu$ l, and less than 50  $\mu$ l will decrease the elution efficiency and the DNA yield.**

7. Store DNA at -20°C or apply to down-stream reactions.